Tetrahedron Letters No. 41, pp 4065 - 4068, 1973. Pergamon Press. Printed in Great Britain.

ROSE OXIDE AND IRIDODIAL FROM <u>Aromia moschata</u> L. (COLEOPTERA : CERAMBYCIDAE)

G. Vidari^{*}, M. De Bernardi, M. Pavan⁺ and L. Ragozzino⁺ Istituto di Chimica Organica and ⁺Istituto di Entomologia Agraria, Università di Pavia, Italy

(Received in UK 29 August 1973; accepted for publication 6 September 1973) Odoriferous substances occur in many Coleoptera and secretions of numerous species have been examined. In most cases the components are thought to function in defence against predators.¹

To our knowledge, no member of the family Cerambycidae has been fully investigated and in this paper we report our studies on the constitution of the liquid secretion from adults of the wide-spread European <u>Aromia moschata</u> L. This Cerambycid lives preferentially on willow-trees during the summer and is well known for its delicate scent.

The secretion is manufactured in thoracic glands and is discharged through an opening on the metasternum, near the articulation of the coxa of the hind legs. It is discharged when the insect is menaced by reptiles, birds etc. and is therefore believed to act as defensive substance.

In earlier studies, the presence of salicylaldehyde² or of a salicylic $e ther^3$ in the secretion has been supposed, yet without any chemical evidence.

On the contrast, in this communication, we report evidence that the secretion of <u>A. moschata</u> L., consists largely of four monoterpenes: <u>cis</u>- and <u>trans</u>rose oxide (1,2), δ - and γ -iridodial (3,4).

Glc examination (2,5% SF-96; temperature programmed from 50° to 170° at 5°/min.) of fresh secretion from one insect, immediately analysed after collection, revealed the presence of the above components in the relative concentrations of 22, 4, 63, 11, respectively.

Stored secretion showed no artefact in glc analysis, even after several weeks. 4

Combined gc-ms analysis showed the same molecular ion at m/e 154 for (1) and (2), corresponding to a possible formula $C_{10}H_{18}O$, and a molecular ion at

* Author to whom correspondence and enquiries should be addressed.



m/e 168, attributed to a likely formula $C_{10}H_{16}O_2$, for (3) and (4). Moreover mass spectra exhibit a similar fragmentation pattern for each of the two pairs of components, suggesting two pairs of stereoisomers. Preparative gas chromatogra phy (2,5% SF-96) allowed to isolate a few milligrams of the main components (1) and (3), each impure of the minor compound.

Infrared spectrum of (1) showed no bands of alcohol or carbonyl functions but a strong band at 885 cm⁻¹ attributed to an epoxydic structure. A trisubst<u>i</u> tuted double bond was demonstrated by absorption at 1680 cm⁻¹ and at 845 cm⁻¹. Pmr spectra showed evident signals only for two methyls on a double bond (3H, δ (CDCl₃) 1.69, J 1 Hz and 3H, δ 1.73, J 1 Hz) and for a secondary methyl group (3H, δ 0.94, J 6 Hz). Mass spectrum gave m/e 139 as base peak, corresponding to the loss of a methyl and indicating a fairly stable cyclic structure. Other significant signals are m/e 41 (43%), 55 (34), 69 (69), 83 (30), 84 (16), 85 (18).

These spectral features suggested for (1) (and likewise for (2)) the structure of the monoterpene rose oxide. Chromatographic and spectroscopical (ir, pmr and mass) comparison with authentic <u>cis</u> and <u>trans</u> isomers of rose oxide, ⁵ confirmed that the more volatile compound is the <u>cis</u> isomer, the less volatile being the <u>trans</u>. Absolute configuration was established for (1) by the negative sign of the rotatory power. Comparison with literature data⁶ showed that (1) is $(-)-(4R)-\underline{cis}$ -rose oxide and that presumably (2) is the (-)-(4R)--trans-isomer.

Components (3) and (4) were identified as isomers of iridodial. Both $\inf rac{a}$ red and pmr data established the presence of aldehyde functions.Losses of CH_3 , CHO, CH_3 CHCHO, along with the presence of a methylcyclopentane ring, were evidenced in the mass spectrum by significant signals at m/e 153, 139, 111-110 and 81. Comparison of the spectral (ir, pmr, ms) data with those of an authentic iridodial⁷ and with previous literature⁸ proved the structure beyond any doubt.

As far as the stereochemistry is concerned, the glc retention times, compared with a mixture of the four stereoisomers⁹ of iridodial from natural sources,¹⁰ allowed to infer that compound (3) is δ -iridodial, whereas the Y-structure could be assigned to (4) only tentatively.

No sensible epimerization seems to occur during the isolation procedure. The other two possible isomers of iridodial were not present in appreciable amount to be detected by glc.

Iridodial has previously been identified in several species of ants,^{1,8} where it has always been found in association with 6-methylhept-5-en-2-one. To our knowledge, among Coleoptera it has been isolated only from the odoriferous glands of <u>Staphylinus olens</u>,¹¹ where it was suggested to have a different defensive role that in ants.

Rose oxide has never been found previously in secretions of Arthropods, whereas it has been isolated from plants (Rosa damascena Mill.,^{12a} Pelargonium graveolens Ait.,^{12a} Ribes nigrum L.,^{12b} Hamanasu oil^{12c}) and to our knowledge represents the first example of a monotcrpene with a tetrahydropyrane ring found in the insect kingdom. It has been reported¹³ that cyclopentanoid deriva tives are formed in insects from mevalonate, presumably by the usual pathways. Whether the compounds present in the secretion of <u>Aromia moschata</u> L. are really synthesized by the insect, or are taken up from plants, resorbed, stored and later used as a defensive scent, as proposed by the earliest authors,^{2,3} is an interesting problem under investigation.

<u>Acknowledgements</u>. The authors thank Sergio Pedrazzini and Alberto Tronconi, technicians of the Institute of Entomologia Agraria, Pavia, for assistance in collecting the insects and obtaining the raw secretion. One of us (G.V.) is indebted to the Ministero della Pubblica Istruzione, Rome, for a research fellowship.

REFERENCES AND FOOTNOTES

- 1. For recent summaries see:
 - (a) J. Weatherston and J.E. Percy in <u>Chemicals controlling Insect Behavior</u>, Edit., M. Beroza, Academic Press, p. 95 (1970),
 - (b) M. Pavan and M. Valcurone Dazzini in <u>Chemical Zoology</u>, Academic Press, vol. 6, p. 365 (1971).
- 2. A.-Ch. Hollande, Ann.Univ.Grenoble Sect. Sci. Med., 21, 459 (1909).
- D.A. Smirnov, <u>Trav.Soc.Imp.Naturalistes St. Pétersbourg, Sect. Zool. Phys.</u>, <u>40</u> (3-4), 1-15 (1911).
- 4. The material was drawn from 170 adults in the summer 1973. The insect were stimolated to spray out the secretion at the top of a test-tube. The liquid was collected in glass capillaries, sealed and stored at -20°.
- 5. Obtained through the courtesy of Prof. Yves-René Naves (Givaudan).
- 6. C.F. Seidel and Coll., <u>Helv.Chim.Acta</u>, <u>44</u>-II (66), 598 (1961).
- 7. We are indebted to Prof. G. Pattenden for sending us the spectra of iridodia from <u>Staphilinus olens</u> (see ref. 11).
- For a review see G.W.K. Cavill in <u>Cyclopentanoid Terpene Derivatives</u>, Edit.,
 W.I. Taylor and A.R. Battersby, Dekker, p. 203 (1969).
- 9. D.J. Mc Gurk and Coll., <u>J.Insect Physiol.</u>, <u>14</u>, 841 (1968).
- 10. We are indebted to Prof. R. Trave (Modena) and to Prof. L. Garanti (Milan) for a generous gift of natural iridodial and for running the reference gas chromatograms.
- 11. S.A. Abou-Donia and Coll., Tetrahedron Letters, 4037 (1971).
- 12. a. G. Ohloff, Fortschr.chem.Forsch., Bd 12/2, 185 (1969).
 - b. E. von Sydow and G. Karlsson, Lebensm-.Wiss.u.Technol., 4 (2), 54 (1971).
 - c. K. Nishimura and Coll., <u>Bull.Chem.Soc.Japan</u>, <u>37</u> (10), 1407 (1964).
- 13. J. Meinwald and Coll., Science, 151, 79 (1966).